THE RENIFORM NEMATODE

J. B. MacGowan

The reniform nematode, <u>Rotylenchulus reniformis</u> Linford & Oliveira 1940, is a severe pathogen of cotton, soybeans, pineapples, and sweet potatoes. It is known to parasitize approximately 160 other plants. It was first observed on the roots of cowpea, <u>Vigna unguiculata</u> (L.) Walp., in the Hawaiian Islands. The reniform nematode was described as a new species in 1940 after 4 years of investigations. The name "reniform" refers to the distinct kidney-shaped appearance of the mature female.

HOSTS:

The reniform nematode is of considerable economic importance. It is distributed around the world and is found in the southern United States. It is one of the most common nematodes found in areas under cultivation in tropical America. It has a lengthy host range which includes many dicots but few monocots. Some of the economically important host plants are:

Banana - Musa x paradisiaca L.

Cabbage - Brassica oleracea L. (Capitata group)

Cantaloupe - Cucumis melo L.

Cassava - Manihot esculenta Crantz

Citrus - Citrus x limonia Osbeck

" - C. maxima (Burm.) Merr.

" - C. sinensis L. Osbeck

Coconut - Cocos nucifera L.

Cotton - Gossypium hirsutum L.

Cowpea - Vigna unguiculata (L.) Walp.

(V. sinensis Endl.)

Clover, crimson - Trifolium incarnatum L.

Eggplant - Solanum melongena L.

Kale - Brassica oleracea L. (Acephala group)

Lettuce - Lactuca sativa L.

Mango - Mangifera indica L.

Okra - Hibiscus esculentus L.

Pigeon pea - Cajanus cajan (L.) Huth
Pineapple - Ananas comosus (L.) Merr.

Pumpkin - Cucurbita moschata (Duchesne)
Poir.

Radish - Raphanus sativus L.

Soybean - Glycine max (L.) Merr.

Sweet potato - Ipomoea batatas (L.) Lam.

Tobacco - Nicotiana tabacum L.

Tomato - Lycopersicon esculentum Mill.

The following plants have been reported as showing immunity or resistance to the reniform nematode:

Barley, wild - <u>Hordeum pusillum Nutt.</u>
Grass, barnyard - <u>Echinochloa crus-galli</u>
(L) Beuv.
Grass, pangola - <u>Digitaria decumbens</u>
Stent.
Mustard - <u>Brassica nigra</u> (L.) W. D. J.
Koch 'Evergreen'
Oats - Avena sativa L. 'Fulghum'

Guava - Psidium guajava L.

Pepper, red-hot - <u>Capsicum annuum</u> L.

Pepper, sweet - <u>Capsicum annuum</u> L.

Sorghum, sweet - <u>Sorghum bicolor</u> (L.)

Moench. (<u>S. vulgare Pers.</u>)

Spinach - <u>Spinacia oleracea</u> L. 'Bloomsdale'

Sugarcane - <u>Saccharum officinarum</u> L.

'C. P. 44-101'

SYMPTOMS AND DAMAGE:

Roots are infected only by young females. Other plant parasitic nematodes become infective during the 2nd larval stage. A general review of the literature indicates that reniform larvae normally do not feed.

The young female enters the root headfirst causing browning and necrosis of damaged cells. She feeds on the phloem cells after puncturing them with her stylet. The phloem cells enlarge 4-6 times their normal size and extend 10-12 cells in all directions from the feeding site. The enlarged phloem cells are not commonly multinucleate nor as large as the giant cells associated with host responses to the feeding of root-knot or cyst nematodes.

In cotton, necrosis of root tissues causes premature decay resulting in dead or unhealthy and stunted plants. Cotton seedlings may harbor as many as 900 egg masses per plant. Reniform nematodes also are associated with incidences of Fusarium wilt as high as 81.4% in wilt susceptible varieties of cotton compared to 10% where reniform nematodes are not present in the soil.

LIFE CYCLE:

The life cycle requires 17-29 days for completion from egg to egg. After the female has deposited her eggs, the developing larvae molt once while still inside the eggs. After hatching, they molt 3 more times to become young adult males or females.

The young females penetrate into the roots about one-third of their body lengths. Some may become completely embedded. The female body enlarges, and approximately 100 eggs are deposited into a transparent quick-hardening substance called the matrix. The matrix is secreted from cells on either side of the vaginal canal during the egg-laying process. Mating males sometimes get caught in the matrix.

BRIEF DESCRIPTION OF GROSS CHARACTERISTICS:

Males, females, and larvae all have: 1) esophageal glands overlapping the intestine; 2) the dorsal gland orifice a distance greater than 1/2 the stylet length posterior to the stylet knobs; and 3) heavily sclerotized lip structures.

The mature female has a swollen kidney-shaped body containing 2 ovaries. Length is 0.38-0.44 Width at vulva is 0.10-0.14 mm. Vulva is located 68-78% of the body length from the head. Stylet and knobs are distinct.

The immature female is eelworm-shaped with 2 ovaries. Length is 0.34-0.42 mm. Vulva is 1ocated 68-73% of the body length from the head. Stylet and knobs are distinct.

The male is eelworm-shaped. Length is 0.34-0.42 mm. Stylet and knobs are reduced. Caudal alae are rudimentary.

IMPORTANCE TO AGRICULTURE:

The reniform nematode has demonstrated that it can adapt and become an aggressively reproducing parasite after 12 generations exposure to an unfavorable host. It is hardy enough to survive in large numbers over long periods of time in fallow soil. Investigations reveal that it can live 6-7 months in stored dry soil. It is a pest with the potential for causing serious damage to economic crops.

SELECTED REFERENCES:

- Ayala, A. 1962. Pathogenicity of the reniform nematode on various hosts. J. Agric. Univ. Puerto Rico. 44:73-82.
- , and C.T. Ramirez. 1964. Host range distribution and bibliography of the reniform nematode, Rotylenchulus reniformis, with special reference to Puerto Rico. J. Agric. Univ. Puerto
- Rico. 48-140-161.
 Birchfield, W. 1962. Host parasite relations of <u>Rotylenchulus reniformis</u> on <u>Gossypium hirsutum</u>. Phytopathology 52:862-865.
- 1972. Differences in host cell responses to the reniform nematode. Phytopathology
- , and L. R. Brister. 1962. New hosts and non-hosts of the reniform nematode. Plant Dis. Reptr. 46:683-685.
- , and J. E. Jones. 1961. Distribution of the reniform nematode in relation to crop failure of cotton in Louisiana. Plant Dis. Reptr. 45:671-673.
- Cohn, E. 1976. Cellular changes induced in roots by two species of the genus Rotylenchulus. Nematologica 22:169-173.
- Dasgupta, D. R., D. J. Raski, and S. A. Sher. 1968. A revision of the genus Rotylenchulus Linford and Oliveira 1940 (Nematoda: Tylenchidae). Proc. Helm. Soc. Wash. 35:169-192.
- Jensen, H. J. 1972. Nematode pests of vegetable and related crops. p. 377-408 in J. M. Webster, ed. Economic nematology. Academic Press, New York.
- Linford, M. B., and J. M. Oliveira. 1940. Rotylenchulus reniformis, nov. gen. n. sp., a nematode parasite of roots. Proc. Helm. Soc. Wash. 7:35-42.
- _, and F. Yap. 1940. Some host plants of the reniform nematode in Hawaii. Helm. Soc. Wash. 7:42-44.
- Nakasono, K. 1973. Studies on post-embryonic development of the reniform nematode. Rotylenchu-<u>lus reniformis</u> Linford and Oliveira (Nematoda: Rotylenchulidae). 1. Morphological changes of the molting larva of a parthenogenetic population. Appl. Ent. Zool. 8:83-96.
- Rebois, R. V. 1973. Effect of soil temperature on infectivity and development of Rotylenchulus reniformis on resistant and susceptible soybeans, Glycine max. J. Nematol. 5:10-13.
- , J. M. Epps, and E. E. Hartwig. 1970. Correlation of resistance Heterodera glycines and Rotylenchulus reniformis. Phytopathology 60:695-700. Correlation of resistance in soybeans to
- Immunity of sugarcane to the reniform nematode. J. Agric. Univ. Puerto Rico 48:162-163.
- Sasser, J. N. 1972. Nematode diseases of cotton. p. 197-214 in J. M. Webster, ed. Economic nema-
- tology. Academic Press, New York.
 Sivakumar, C. V., and A. R. Seshadri. 1971. Life history of the reniform nematode, Rotylenchulus reniformis Linford and Oliveira, 1940. Indian J. Nematol. 1:7-20.
- . 1972. Histopathology of infection by the reniform nematode, Rotylenchulus reniformis Linford and Oliveira, 1940 on castor, papaya and tomato. Indian J. Nematol. 2:173-181.